

Triterpenoids from *Abies* species

23.* A lanostane analog of cytotoxic cycloartanolide and other lactones from needle-free shoots of Siberian fir

T. P. Kukina, M. M. Shakirov, and V. A. Raldugin*

*Novosibirsk Institute of Organic Chemistry, Siberian Branch of the Russian Academy of Sciences,
9 prosp. Akad. Lavrent'eva, 630090 Novosibirsk, Russian Federation.
Fax: +7 (383 2) 35 4752. E-mail: raldugin@nioch.nsc.ru*

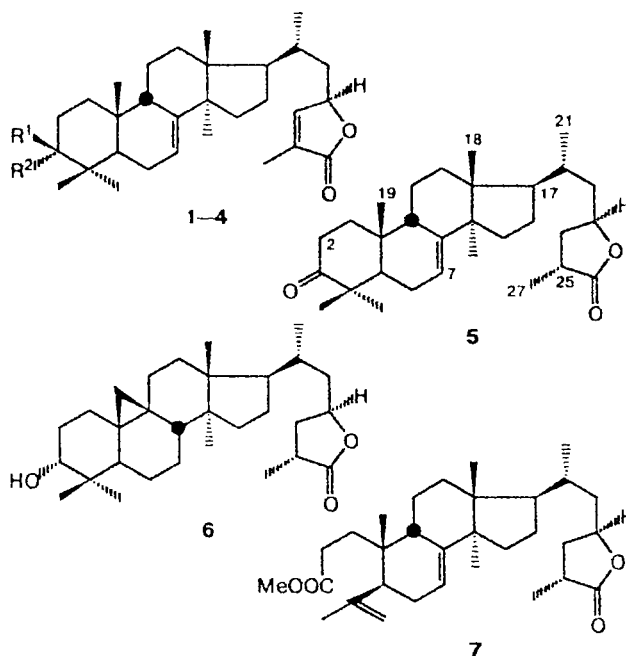
Two new lanostane lactones were isolated from a neutral part of an ethereal extract of dried needle-free shoots of Siberian fir. Their structures were proposed based on comparison of their ^{13}C and ^1H NMR spectra with the spectra of known triterpenoid lactones with similar structures.

Key words: triterpenoids; Siberian fir; lactones; NMR spectra.

The bark and coniferous needles of Siberian fir (*Abies sibirica* Ledeb.) contain triterpenoid lactones, the best known of which are abieslactone (1)^{2–4} and its 3-oxo- and 3-hydroxy-analogs 2 and 3, respectively.⁴ For lactone 1 itself,² unlike its derivatives,⁵ no clear-cut biological activity has been discovered so far.

We studied the chemical composition of an ethereal extract of needle-free 1–3-years old shoots of Siberian fir, which are present in the commercial "fir green." It was found that these shoots contain lactones 1–3 and some other hitherto unknown lactones including natural acetate 4. An oxo lactone with a melting point of 187–188 °C was identified as compound 5 based on the data of ^{13}C NMR (Table 1) and ^1H NMR spectra (see Experimental) and on comparison of these spectra with those of closely related analogs, viz., natural oxo lactone 2⁶ and hydroxy lactone 6⁷. The stereochemistry of this compound corresponds to that of other triterpenoids from Siberian fir, viz., oxolactone 2 and methyl abiesolidate 7,⁸ whose structures were established previously by X-ray diffraction analysis.

The second lactone (m.p. 238–240 °C) has the molecular formula $\text{C}_{30}\text{H}_{42}\text{O}_4$ (determined by high-resolution mass spectrometry). Its ^{13}C NMR spectrum (see Table 1) can be represented as a combination of the corresponding spectra of the steroid part of lactone 2 (see Ref. 6) and the spiropyranolactone fragment of the molecule of pseudolarolid B (8),⁹ which is a cytotoxic cycloartanolide from the extract of seeds of a Chinese plant, *Pseudolarix kaempferi* Gord. (the Pinaceae Lindl. family). Similar correspondence can also be noted for the ^1H NMR spectra of lactone 8 and the compound in question (Table 2 and Experimental); hence, structure 9 was proposed for this compound.

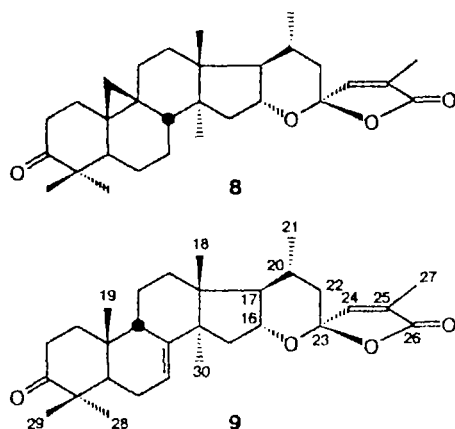


1: $\text{R}^1 = \text{H}$; $\text{R}^2 = \text{OMe}$ 3: $\text{R}^1 = \text{H}$; $\text{R}^2 = \text{OH}$
2: $\text{R}^1 + \text{R}^2 = \text{O}$ 4: $\text{R}^1 = \text{H}$; $\text{R}^2 = \text{OAc}$

The ^1H NMR spectrum is more sensitive to the appearance of a pyran ring fused with ring D of the steroid nucleus in the molecule of lactone 9. The shape and position of the signal for the H(7) proton correspond to the values expected¹⁰ for 3-oxo-9 β -lanost-7-enoids, and the signals for the $\text{H}_2\text{C}(2)$ protons, which almost coincide in the spectra of oxo lactones 2 and 5, are substantially different in the ^1H NMR spectrum of lactone 9.

The scarcity of compound 9 at our disposal precluded its X-ray diffraction analysis needed to establish unambiguously its structure.

* For Part 22, see Ref. 1.

**Table 1.** Data of the ^{13}C NMR spectra (δ) of compounds 2, 5, 6, 8, and 9 (CDCl_3)

Atom	2	5	6	8	9
C(1)	34.20 (t)	34.13 (t)			33.30 (t)
C(2)	34.28 (t)	34.29 (t)			33.55 (t)
C(3)	218.92 (s)	218.61 (s)			219.14 (s)
C(4)	47.01 (s)	46.91 (s)			46.76 (s)
C(5)	52.35 (d)	52.27 (d)			53.16 (d)
C(6)	23.01 (t)	22.94 (t)			22.62 (t)
C(7)	121.64 (d)	121.54 (d)			122.16 (d)
C(8)	148.48 (d)	148.43 (s)			146.86 (s)
C(9)	45.67 (d)	45.44 (d)			44.67 (d)
C(10)	35.80 (s)	35.72 (s)			35.91 (s)
C(11)	20.88 (t)	20.82 (t)			19.13 (t)
C(12)	34.39 (t)	34.29 (t)			34.02 (t)
C(13)	44.15 (s)	44.03 (s)		44.6 (s)	43.42 (s)
C(14)	51.93 (s)	51.87 (s)		46.2 (s)	49.31 (s)
C(15)	33.01 (t)	32.98 (t)		43.4 (t)	41.11 (t)
C(16)	28.21 (t)	28.23 (t)		75.5 (d)	75.45 (d)
C(17)	53.47 (d)	53.45 (d)		55.6 (d)	56.30 (d)
C(18)	22.51 (q)	22.44 (q)			22.98 (q)
C(19)	23.12 (q)	23.06 (q)			24.52 (q)
C(20)	33.49 (d)	33.09 (d)	33.06	25.4 (d)	25.36 (d)
C(21)	18.38 (q)	18.14 (q)	18.12	20.2 (q)	20.14 (q)
C(22)		42.53 (t)	42.76	37.8 (t)	37.61 (t)
C(23)		75.77 (d)	76.03	107.2 (s)	107.09 (s)
C(24)		36.37 (d)	36.44	146.3 (d)	146.00 (d)
C(25)		34.07 (d)	34.17	130.7 (s)	130.74 (s)
C(26)		179.91 (s)	180.24	172.3 (s)	172.10 (s)
C(27)		15.84 (q)	15.91	10.4 (q)	10.25 (q)
C(28)	21.30 (q)	21.24 (q)			20.73 (q)
C(29)	28.00 (q)	27.87 (q)			28.39 (q)
C(30)	27.41 (q)	27.34 (q)			26.35 (q)

Experimental

Melting points were determined on a Koffler hot-stage apparatus. IR spectra were recorded on a UR-20 instrument. NMR spectra were measured for CDCl_3 solutions on Bruker DRX-500 (operating at 500.13 MHz for ^1H and at 125.76 MHz for ^{13}C) and Bruker AM-400 (operating at 400.13 MHz for ^1H) spectrometers. Mass spectra were run on a Finnigan MAT 8200 instrument (EI, direct inlet, $T_{\text{source}} =$

Table 2. Data of the ^1H NMR spectra of compounds 8⁹ and 9 (CDCl_3 , tetramethylsilane)

Proton	δ (J/Hz)	
	lactone 8 ⁹	lactone 9
HC(15 α)	1.42 (dd)	**
HC(15 β)	1.96 (dd, $J = 13, 7.5$)	2.04 (dd, $J = 12.0, 8.0$)
HC(16)	4.43 (q, $J = 7.5$)	4.49 (q, $J = 7.5$)
H ₃ C(21)	0.99 (d, $J = 5.5$)	1.01 (d, $J = 5.5$)
HC(24)	6.83 (q, $J = 1$)*	6.82 (q, $J = 1.5$)
H ₃ C(27)	1.87 (d, $J = 1$)	1.87 (d, $J = 1.5$)

* In Ref. 9, this signal was named a doublet.

** The signal was overlapped by other signals.

260 °C, 70 eV). Optical rotation was determined in CHCl_3 on a Polamat A polarimeter at 580 nm.

Column chromatography was carried out on KSK SiO₂; the ratio of compound to sorbent was ~1 : 20; hexane–ether (10 to 25%) was used as the eluent. TLC was carried out on Silufol plates; compounds were visualized by spraying with concentrated H_2SO_4 .

Fir bough was gathered in winter 1995 at the territory of the Botanical Garden of the Siberian Branch of the RAS (Novosibirsk); shoots were separated from needles, dried at room temperature, and ground to a particle size of less than $1 \times 1 \times 1$ mm.

Isolation of the lactone-containing fraction from the extract of shoots. Ground shoots (360 g) were extracted for 113 h at 20 °C with a mixture of ether and ethyl acetate (1 : 1). The extract was decanted, filtered, and treated successively with a 2% aqueous solution of NaOH (2×300 mL) and water (2×200 mL). Drying and removal of the solvent gave 19.6 g of total neutral components of the extract, which was distributed between 100 mL of 80% aqueous ethanol and 150 mL of *n*-pentane. The lower layer (aqueous ethanol) was separated, and the pentane layer was washed with aqueous ethanol (2×100 mL). The aqueous-ethanolic extracts were combined, washed with pentane (2×100 mL), diluted with an equal volume of brine and extracted with a 1 : 1 ether–ethyl acetate mixture (2×100 mL). Drying of the extract (Na_2SO_4) and evaporation of the solvent gave 2.95 g of a lactone-containing fraction as a yellowish oil in a yield of 15% of total of neutral compounds.

Isolation of lactones 4, 5, and 9. Chromatography of the resulting fraction (2.95 g) gave successively a mixture of mono- and diterpenoid nonlactone compounds (0.81 g), a mixture of nonpolar lactones and accompanying compounds (fraction A, 0.57 g), and a fraction of more polar substances (1.24 g). Repeated chromatography of fraction A gave a mixture of lactones (0.11 g) containing lactone 1, acetate 4 (0.06 g) identical with an authentic sample⁴ according to TLC and ^1H NMR spectrum, a mixture (0.07 g) of β -sitosterol and compounds 5 and 9 (fraction B), oxo lactone 2 (0.18 g), and a mixture (0.15 g) of hydroxy lactone 3 and unidentified substances. Fraction B was treated with $\text{Ac}_2\text{O}/\text{Py}$ under known conditions,⁴ and subsequent chromatography gave β -sitosterol acetate (0.03 g), lactone 5 (0.01 g), and lactone 9 (0.01 g).

(23*R*,25*R*)-3-Oxolanost-7-en-26,23-olide (5). Crystals with m.p. 187–188 °C (ether–pentane), $[\alpha]_{\text{D}}^{20} + 17^\circ$ (c 0.59). IR (CCl_4), ν/cm^{-1} : 1770 (γ -lactone), 1703 ($\text{C}=\text{O}$), 1380, 1360, 1170. MS, m/z (I_{rel} (%)): 454.34540 $[\text{M}]^+$ (21), 439 (100), 421 (15), 133 (16), 121 (18), 107 (17), 95 (22). Calculated for $\text{C}_{30}\text{H}_{46}\text{O}_5$: 454.34467. ^1H NMR, δ : 0.78, 0.99,

1.01, 1.07, 1.08 (all s, 5 Me); 0.93 (d, 3 H, C(21)H₃, $^3J_{21,20} = 6.5$ Hz); 1.27 (d, 3 H, C(27)H₃, $^3J_{27,25} = 7.0$ Hz); 2.47 (dd, 2 H, C(2)H₂, $J = 6.0$ and 8.5 Hz); 2.69 (sextet, 1 H, H(25), $^3J_{25,24} = ^3J_{25,27} = 7.5$ Hz); 4.63 (m, 1 H, H(23)); 5.62 (dd, 1 H, H(7), $J = 7.5$ and 3.0 Hz). The data of the ^{13}C NMR spectrum are given in Table 1.

(16R,23S)-3-Oxo-16,23-epoxyabnosta-7,24-dien-26,23-olide (9). Colorless crystals, m.p. 238–240 °C (ether–pentane), $[\alpha]_{580}^{20} +26.1^\circ$ (c 0.65). IR (CHCl₃), ν/cm^{-1} : 1770 (γ -lactone), 1705 (C=O), 1390, 1330, 980, 940, 860. MS, m/z (I_{rel} (%)): 466.30855 [$\text{M}]^+$ (34), 451(78), 433(11), 428(14), 323(53), 313(13), 207(75), 137(100), 119(27). Calculated for C₃₀H₄₂O₄: 466.30829. ^1H NMR (CDCl₃), δ : 0.87, 0.93, 1.00, 1.07, 1.08 (all s, 5 Me); 1.01 (d, 3 H, H₃C(21), $^3J_{21,20} = 6$ Hz); 1.42 (dd, 1 H, HC(5), $J = 12.0$; 2.5 Hz); 2.22 (m, 1 H, HC(9)); 2.41 (ddd, 1 H, H^aC(2), $J = 15.5, 8.5, 6.0$ Hz); 2.55 (ddd, 1 H, H^bC(2), $J = 15.5, 10.0, 4.0$ Hz); 5.66 (dt, 1 H, HC(7), $J = 7.5, 3.0$ Hz). The signals corresponding to signals of lactone **8** are listed in Table 2.

References

1. V. V. Grishko, A. G. Druganov, M. M. Shakirov, and V. A. Raldugin, *Izv. Akad. Nauk, Ser. Khim.*, 1998, 519 [*Russ. Chem. Bull.*, 1998, **47**, 502 (Engl. Transl.)].
2. S. Uyco, J. Okada, S. Matsunaga, and J. W. Rowe, *Tetrahedron*, 1968, **24**, 2859.
3. J. P. Kutney, W. D. Westcott, F. H. Allen, N. W. Isaacs, O. Kennard, and W. D. S. Motherwell, *Tetrahedron Lett.*, 1971, 3463.
4. N. I. Yaroshenko and V. A. Raldugin, *Khim. Prirod. Soedin.*, 1989, 220 [*Chem. Nat. Compd.*, 1989, 188 (Engl. Transl.)].
5. J. Takayashi, R. Tanaka, S. Matsunaga, H. Ueyama, H. Tokuda, T. Hasegawa, A. Nishino, and A. Iwashima, *Cancer Lett.*, 1990, **53**, 141.
6. R. Tanaka and S. Matsunaga, *Phytochemistry*, 1991, **30**, 1983.
7. A. F. Barrero, J. F. Sanchez, E. J. A. Manzaneda, M. Munoz, and A. Hardour, *Phytochemistry*, 1992, **31**, 615.
8. V. A. Raldugin, Yu. V. Gatilov, T. V. Rybalova, and Ya. V. Rashkes, *Khim. Prirod. Soedin.*, 1986, 688 [*Chem. Nat. Compd.*, 1986 (Engl. Transl.)].
9. G.-F. Chen, Z.-L. Li, De-Ji Pan, G.-M. Tang, X. He, G.-Y. Xu, K. Chen, and K.-H. Lee, *J. Nat. Prod.*, 1993, **56**, 1114.
10. V. A. Raldugin, and S. A. Shevtsov, *Khim. Prirod. Soedin.*, 1990, 443 [*Chem. Nat. Compd.*, 1990, 373 (Engl. Transl.)].

Received January 12, 1998;
in revised form March 24, 1998